Large-scale inversions between human reference assemblies

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Abstract

The detection of large-scale inversions between intra-species genomes is fundamental to understand the dynamics of chromosomal evolution, namely to identify hallmarks associated with diseases. In this paper, we explore a method that finds exact or approximate large-scale inversions, for regions larger than 500 kb, between the three most popular assembled human genomes. We identify the relative positions for each chromosome. The inversions are computed in two maps showing at a glance the associated regions.

Introduction

How genome architecture and which macroevolutionary events emerge through time are fundamental to understand the dynamics of species evolution, namely their origin and speciation patterns [3]. Several insights into chromosome evolution have been traditionally achieved by cytogenetic procedures, for example G-banding, and by molecular karyotyping approaches, such as fluorescence in situ hybridization (FISH). More recently, array-based methods became very popular [2].

Advances in sequencing technology have increased the number of digital human genomes, raising conditions towards intra-species characteristics and diversity research. Consequently, computational approaches emerged [5], bringing high resolution, accuracy and speed with less costs. However, the de novo assembly of the next generation sequencing (NGS) reads is still problematic, mainly because the alignment of the reads from these new genomes to a high quality reference genome remains a critical aspect of data interpretation. Nevertheless, the human reference assembly is the highest quality mammalian assembly available. The main reference genome assemblies are those from the Genome Reference Consortium (GRC 38) [1], the J. Craig Venter Institute (HuRef) [4] and the Washington U. School of Medicine (CHM 1.1).

In this paper, we detect the large-scale (larger than 500 Kb) inversions between these three reference genome assemblies, exploring fundaments from an unsupervised alignment-free method [9], that is based in the hability to efficiently model the repetitiveness of genomic sequences [8].

Method

The method involves the estimation of the amount of conditional exclusive information, i. e., using only information from a reference that is required to represent a certain region of the sequence or sub-sequence [6]. The process is as follows:

- 1. Convert symbols outside $\mathcal{A} = \{A, C, G, T, N\}$ into "N";
- 2. Pseudo-randomize the "N" symbols (with uniform distribution);
- 3. Invert the first sequence;
- 4. Load to the FCM [7] the sequence created in step 3;
- 5. Compress the second sequence using the FCM created in step 4;
- 6. Filter the information sequence generated in 5;
- 7. Use a threshold of 1.6 to segment the sub-sequence regions;
- 8. Paint sub-sequence regions with different colors;
- 9. For each sub-sequence region, repeat the process from step 3 to 8, using the second sequence as target;

The colors are calculated automatically using a HSV scheme where only V varies.

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Results

For the results presented in this section we have downloaded the genomes from the NCBI¹ site. Chromosome Y from CHM genome is absent and thus we have not reported results associated with this sequence.

Fig. 1 shows the maps from the large-scale inversions between A (GRC) and B (HuRef), while Fig. 2 shows the inversions between A(GRC) and C (CHM) assemblies. In respect to A/B, there are inversions in chromosomes 1, 2, 7, 9, 10, 11, 15 and Y. Specifically to chromosome 1, the inversions are contained in the pericentric regions (around 119 Mb). This region is also inverted between human and chimpanzee species, although in a much larger density [9].

On the other hand, for A/C the inversions are present between chromosomes 1, 2, 5, 7, 8, 9, 10, 11, 14, 15, 16, 17, 22, X. From these, most of the inversions are contained in pericentric regions, a major factor of dynamism across individuals of the same species.

Conclusions

We have proposed a procedure to detect exact or approximate inversions, larger than 500 Kb, between reference assembled genomes. The inversions have been computed in a information map.

Besides specific characteristics, the reference assembly from the Washington U. School of Medicine (CHM) seems to have a higher number of inversions compared with the Genome Reference Consortium (GRC 38), than with the J. Craig Venter Institute (HuRef).

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¹ftp://ftp.ncbi.nlm.nih.gov/genomes/Homo_sapiens/Assembled_ chromosomes/seq/



Figure 1: Large-scale inversions between GRC (A) and HuRef (B) assemblies for each chromosome. The information maps show exact or approximate inversions with length higher than 500 kb. Each position associated with inversions, in the HuRef chromosomes, is reported in the table and marked with a greek letter according to the map.



Figure 2: Large-scale inversions between GRC (A) and CHM (C) assemblies for each chromosome. The information maps show exact or approximate inversions with length higher than 500 kb. Each position associated with inversions, in the CHM chromosomes, is reported in the table and marked with a greek letter according to the map.